

# Package: qp (via r-universe)

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**Repository** <https://kaiaragaki.r-universe.dev>

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<b>absorbances</b>	<i>Absorbances from a protein quantification</i>
--------------------	--

---

## Description

Absorbances from a BCA protein quantification in a `data.frame`

## Usage

`absorbances`

## Format

`absorbances`:

A `data.frame` with 96 rows and 5 columns::

**.row** The row of the 96 well plate, where 1 refers to the top row.

**.col** The column of the 96 well plate, where 1 refers to the left column.

**.abs** The absorbance of the contents of the well at 562nm.

**sample\_type** Denotes whether the sample is a standard or an unknown (sample).

**index** Denotes individual standards/samples, where each gets its own index.

---

**abs\_to\_col***Convert an absorbance to a hexidecimal color*

---

**Description**

Takes an absorbance and converts it to a hexidecimal color. For the default qp\_pal palette, this should provide a color that approximates real life color at the given absorbance.

**Usage**

```
abs_to_col(abs, pal)
```

**Arguments**

abs	Numeric. Absorbances.
pal	Character. A vector of hexidecimal colors.

**Details**

The absorbances have typical baseline absorbance (~ 0.07) removed, and then an index is calculated with a logistic curve of maximum 100 and a center of 0.15.

**Value**

Character. Hexidecimal colors corresponding to absorbances.

---

**dilute***Calculate dilution from known concentrations*

---

**Description**

Calculate dilution from known concentrations

**Usage**

```
dilute(c1, c2 = min(c1), v2, round_for_pipettes = TRUE)
```

**Arguments**

c1	Numeric. Initial concentration of sample.
c2	Numeric. Target concentration of sample.
v2	Numeric. Target final volume of sample. If <code>round_for_pipettes = TRUE</code> , assumes volume is uL.
round_for_pipettes	Logical. If TRUE, rounds values to the accuracy of standard pipettes using <code>make_pipette_vol</code> .

**Value**

a data.frame, with `sample_to_add` as the volume of sample to add, and `add_to` as the volume to dilute the sample into.

**Examples**

```
dilute(203, 70, 10)
dilute(203, 70, 10, round_for_pipettes = FALSE)
# Vectorized:
dilute(c(8, 10, 12), c(4, 5, 6), c(7, 8, 9))
```

<code>make_pipette_vol</code>	<i>Round volume to be pipette-compatible</i>
-------------------------------	--

**Description**

Round volume to be pipette-compatible

**Usage**

```
make_pipette_vol(x)
```

**Arguments**

<code>x</code>	Numeric. Volume to be rounded
----------------	-------------------------------

**Value**

Numeric. Rounded volume.

**Examples**

```
make_pipette_vol(104.13398)
make_pipette_vol(15.3331)
make_pipette_vol(9.9211)
# Vectorized:
make_pipette_vol(c(104.13398, 15.3331, 9.9211, NA, -100.1))
```

---

**qp***Quantify protein concentration from a MicroBCA assay*

---

## Description

Quantify protein concentration from a MicroBCA assay

## Usage

```
qp(  
  x,  
  replicate_orientation = c("h", "v"),  
  sample_names = NULL,  
  remove_empty = TRUE,  
  ignore_outliers = c("all", "samples", "standards", "none"),  
  standard_scale = c(0, 2^(2:7) - 5),  
  n_replicates = 3,  
  wavelength = 562  
)
```

## Arguments

**x** A spectramax, gp, or data.frame object, or path to SPECTRAmax .xls(x)/.txt file.

**replicate\_orientation** Either 'h' or 'v' - see Details.

**sample\_names** Optional character vector of sample names.

**remove\_empty** Should wells that have less absorbance than the lowest standard be dropped?

**ignore\_outliers** Character. From which group - samples or standards - should outliers be detected and removed?

**standard\_scale** Numeric. Known concentrations of standards, in the order they appear.

**n\_replicates** Numeric. The number of technical replicates.

**wavelength** Numeric. The wavelength absorbance was captured.

## Details

If x is a spectramax, the standards must start in the upper left corner in the order dictated by standard\_scale. Whether this is from left to right or top to bottom can be specified in replicate\_orientation. Note that replicate\_orientation specified the direction that REPLICATES lie, NOT the direction the samples flow (which will be perpendicular to the replicates).

Note: replicate\_orientation, n\_replicates, and wavelength will be silently ignored if x is not a spectramax or path to a spectramax

**Value**

a tibble

**Examples**

```
data <- system.file("extdata", "absorbances.txt", package = "qp")
qp(data, replicate_orientation = "h")
```

*qp\_add\_names*

*Add sample names*

**Description**

Add sample names

**Usage**

```
qp_add_names(x, ...)
## S3 method for class 'list'
qp_add_names(x, sample_names = NULL, ...)
## S3 method for class 'data.frame'
qp_add_names(x, sample_names = NULL, ...)
```

**Arguments**

- x A `data.frame` (or a list containing one) that contains columns `index` (which denotes sample number) and `sample_type`, which should be either "unknown" or "standard".
- ... Unused
- sample\_names Optional character vector. If `NULL`, uses sample index. In a standard workflow, the index is the order the sample appears in the plate

**Examples**

```
df <- expand.grid(
  index = c(1, 1, 2, 2, 2, 3),
  sample_type = c("standard", "unknown")
)
df

# You don't get to name standards:
qp_add_names(df, c("a", "b", "c"))

# If there aren't enough names, will use index
```

```
qp_add_names(df, c("a", "b"))

# No names provided will use index by default
qp_add_names(df)
```

---

**qp\_add\_std\_conc***Add known concentrations of protein to standard samples*

---

**Description**

Add known concentrations of protein to standard samples

**Usage**

```
qp_add_std_conc(x, standard_scale = c(0, 2^((2:7) - 5)), ...)

## S3 method for class 'data.frame'
qp_add_std_conc(x, standard_scale = c(0, 2^((2:7) - 5)), ...)

## S3 method for class 'list'
qp_add_std_conc(x, standard_scale = c(0, 2^((2:7) - 5)), ...)
```

**Arguments**

**x** A `data.frame` containing a `sample_type` and `index` columns. See details.

**standard\_scale** A numeric vector giving the concentrations of the standards. The units are arbitrary, but will determine the units of the output concentrations.

**...** Unused

**Details**

Input is expected to have two columns:

- `sample_type`: A character vector denoting which samples are standards with "standard". All other values will be considered unknowns.
- `index`: A numeric column denoting the sample number. Index 1 will correspond to the first item in `standard_scale`, 2 will be the second, etc.

**Value**

Same type as `x`, with a `.conc` column

## Examples

```

abs <- expand.grid(
  sample_type = c("standard", "unknown"),
  index = 1:7
)
abs

qp_add_std_conc(abs)

# Can add custom scale - doesn't have to be 'in order' or unique:
qp_add_std_conc(abs, c(1, 4, 2, 2, 3, 0.125, 7))

# Will warn - more values in `standard_scale` than standard indices
# Will drop extra
qp_add_std_conc(abs, 1:8)

# Will error - fewer values in `standard_scale` than standard indices
if (FALSE) {
  qp_add_std_conc(abs, 1:6)
}

```

*qp\_calc\_abs\_mean*

*Calculate absorbance means with optional outlier removal*

## Description

Calculate absorbance means with optional outlier removal

## Usage

```

qp_calc_abs_mean(x, ignore_outliers = c("all", "standards", "samples", "none"))

## S3 method for class 'data.frame'
qp_calc_abs_mean(x, ignore_outliers = c("all", "standards", "samples", "none"))

## S3 method for class 'list'
qp_calc_abs_mean(x, ignore_outliers = c("all", "standards", "samples", "none"))

```

## Arguments

- x A `data.frame` or `list` containing a `data.frame` named `qp`. See details.
- ignore\_outliers Which sample types should have outliers ignored from their mean calculations?  
If `.is_outlier` column is supplied, this argument is ignored.

## Details

Input data `.frame` must contain the following columns:

- `sample_type`. Character. Must contain values either "standard" or "unknown"
- `index`. Numeric. Denotes sample number.
- `.abs`. Numeric. Contains absorbance values.
- If a boolean `.is_outlier` is supplied, that will be used instead.

## Value

The input tibble with an `.is_outlier` column and a `.mean` column

## Examples

```
library(dplyr)

abs <- expand.grid(
  sample_type = c("standard", "unknown"),
  index = 1:7,
  rep = 1:3
) |>
  dplyr::arrange(sample_type, index, rep)

abs$.abs <- abs(rnorm(nrow(abs), mean = abs$index))

# Selecting different subsets for outlier removal
qp_calc_abs_mean(abs, "none")

qp_calc_abs_mean(abs, "standards")

qp_calc_abs_mean(abs, "samples")

qp_calc_abs_mean(abs, "all")

# If an `is_outlier` column is provided, that will be used instead:
abs$.is_outlier <- rep(c(TRUE, FALSE), length.out = nrow(abs))

qp_calc_abs_mean(abs)
```

## Description

Predict concentrations from standards fit

**Usage**

```
qp_calc_conc(x, ignore_outliers = TRUE, group_cols = c("sample_type", "index"))
```

**Arguments**

- x A list. See details.
- ignore\_outliers Boolean. Should outliers be considered when calculating the mean? See details.
- group\_cols Character vector. Columns to group by before taking the mean.

**Details**

The supplied list should contain two items - fit, generated by qp\_fit, and qp, a data.frame. qp should contain the following:

- Columns used in fit. Usually, this is .log2\_abs
- Any columns in group\_cols
- If ignore\_outliers = TRUE, .is\_outlier will be used if supplied, or created if not.

**Value**

Returns a list with the input fit and data.frame, with additional columns:

- .pred: The predicted value from the provided model
- .pred\_conc: .pred, transformed by conc\_transform
- .pred\_conc\_mean: The mean of .pred\_conc, sans samples where column .is\_outlier == TRUE

**Examples**

```
data <- system.file("extdata", "absorbances.txt", package = "qp")
calculated <- qp(data, replicate_orientation = "h")

# Making a minimal object:
calculated$qp <- calculated$qp |>
  dplyr::select(
    .log2_abs, sample_type, index, .is_outlier
  )

calculated

qp_calc_conc(calculated)
```

---

qp\_dilute                    *Calculate dilutions from predicted concentrations*

---

### Description

Calculate dilutions from predicted concentrations

### Usage

```
qp_dilute(x, ...)

## S3 method for class 'data.frame'
qp_dilute(
  x,
  target_conc = NULL,
  target_vol = 15,
  remove_standards = FALSE,
  pipette_vol_compat = TRUE,
  ...
)

## S3 method for class 'list'
qp_dilute(
  x,
  target_conc = NULL,
  target_vol = 15,
  remove_standards = FALSE,
  ...
)
```

### Arguments

x	A data.frame or list containing a data.frame named qp with a column named .pred_conc or .pred_conc_mean. If both, will favor .pred_conc_mean.
...	Unused
target_conc	Numeric vector. Target concentration in (mg/mL) protein. If length == 1, recycled.
target_vol	Target volume in uL. If length == 1, recycled.
remove_standards	Boolean. Should standards be removed from results?
pipette_vol_compat	Boolean. Should returned numbers be rounded to the typically precision of a pipette?

### Value

Same as input, with the volumes of lysate and volumes of diluent to add.

## Examples

```
df <- data.frame(.pred_conc = 1)
qp_dilute(df, target_conc = 0.5, target_vol = 30)

# Many sample and target concentrations
df2 <- data.frame(.pred_conc = 1:3)
qp_dilute(df2, target_conc = c(0.1, 0.4, 0.8), target_vol = 30)

# Takes a list, so long as it has a data.frame named qp as one of the items:
ls <- list(qp = data.frame(.pred_conc = 3))
qp_dilute(ls, target_conc = 0.5, target_vol = 30)
```

**qp\_fit**

*Fit an lm using standards absorbances*

## Description

Fit an lm using standards absorbances

## Usage

```
qp_fit(x)

## S3 method for class 'data.frame'
qp_fit(x)

## S3 method for class 'list'
qp_fit(x)
```

## Arguments

**x** A data.frame or list containing a data.frame under the name qp. See details.

## Details

The supplied data.frame must have the following columns:

- **sample\_type**. Character. If not 'standard', assumed to be a sample
- **.is\_outlier**. Boolean. If TRUE, assumed to be outlier and removed from fitting. If FALSE or NA, used for fitting. If unsupplied, will create one with all values set to NA.
- **.conc**. Numeric. Known concentration of standard.
- **.log2\_abs**. Numeric. The log2 of the absorbances

**Value**

A list containing:

- `fit`, an `lm` object fit with the formula `.log2_conc ~ .log2_abs`, fit using non-outlier standards
- `qp`, the input data

**Examples**

```
absorbances |>
  qp_add_std_conc() |>
  qp_fit()
```

qp_mark_outliers	<i>Mark absorbance outliers</i>
------------------	---------------------------------

**Description**

Mark absorbance outliers

**Usage**

```
qp_mark_outliers(x, ignore_outliers = c("all", "standards", "samples", "none"))

## S3 method for class 'data.frame'
qp_mark_outliers(x, ignore_outliers = c("all", "standards", "samples", "none"))

## S3 method for class 'list'
qp_mark_outliers(x, ignore_outliers = c("all", "standards", "samples", "none"))
```

**Arguments**

<code>x</code>	A <code>data.frame</code> or <code>list</code> containing a <code>data.frame</code> named <code>qp</code> . See details.
<code>ignore_outliers</code>	Which sample types should have outliers marked?

**Details**

Input `data.frame` must contain the following columns:

- `sample_type`. Character. Must contain values either "standard" or "unknown"
- `index`. Numeric. Denotes sample number.
- `.abs`. Numeric. Contains absorbance values.

**Value**

The input `tibble` with an `.is_outlier` column

## Examples

```
df <- data.frame(
  sample_type = rep(c("standard", "unknown"), each = 3),
  index = c(1, 1, 1, 2, 2, 2),
  .abs = c(1, 1, 1, 1, 1, 2)
)

qp_mark_outliers(df, ignore_outliers = "all")
qp_mark_outliers(df, ignore_outliers = "standards")
qp_mark_outliers(df, ignore_outliers = "samples")
qp_mark_outliers(df, ignore_outliers = "none")
```

**qp\_pal**

*The default color palette for qp*

## Description

It attempts to match the real life colors of a protein quantification experiment, in combination with `abs_to_col`

## Usage

`qp_pal`

## Format

An object of class `character` of length 100.

**qp\_plot\_plate**

*View the absorbances of an analyzed qp as they were on the plate*

## Description

View the absorbances of an analyzed qp as they were on the plate

## Usage

`qp_plot_plate(x, size = 15)`

## Arguments

- |                   |  |
|-------------------|--|
| <code>x</code>    | A <code>data.frame</code> with <code>.row</code> , <code>.col</code> , and <code>.abs</code> columns |
| <code>size</code> | The size of the points used to illustrate the wells. Passed to <code>geom_point</code> .             |

**Value**

```
a ggplot
```

**Examples**

```
qp_plot_plate(absorbances)
```

---

qp\_plot\_standards      *View an absorbance/concentration plot*

---

**Description**

View an absorbance/concentration plot

**Usage**

```
qp_plot_standards(x)
```

**Arguments**

x                  The output of qp or qp\_calc\_conc

**Value**

```
a ggplot
```

**Examples**

```
absorbances |>
  qp() |>
  qp_plot_standards()
```

---

qp\_remove\_empty      *Remove empty wells from data*

---

**Description**

Remove empty wells from data

**Usage**

```
qp_remove_empty(x)
```

```
## S3 method for class 'data.frame'
qp_remove_empty(x)
```

```
## S3 method for class 'list'
qp_remove_empty(x)
```

### Arguments

- x A `data.frame` or `list` containing a `data.frame` named `qp` containing columns `.pred_conc` and `sample_type`. See details.

### Details

This function keeps any columns with positive `.pred_conc` or `sample_type == "standard"`

### Value

Same as input

### Examples

```
df <- expand.grid(
  .pred_conc = 0:1,
  sample_type = c("standard", "unknown")
)
df
qp_remove_empty(df)
```

`qp_report`

*Create a report for a protein quantification experiment*

### Description

Create a report for a protein quantification experiment

### Usage

```
qp_report(qp, output_file, other = list())
```

### Arguments

- |                          |   |
|--------------------------|---|
| <code>qp</code>          | Likely the output from <code>qp</code> AND <code>qp_dilute</code> .   |
| <code>output_file</code> | Character. The path of the file to export, including <code>.html</code>   |
| <code>other</code>       | Generally used for Shiny application. Assumes a named list of key-values that will be used to document report parameters. |

## Examples

```
## Not run:  
absorbances |>  
  qp() |>  
  qp_dilute() |>  
  qp_report(  
    "~/my_report.html",  
    other = list(key = "value") # Essentially metadata  
)  
  
## End(Not run)
```

---

qp\_summarize

*Summarize output from qp pipeline*

---

## Description

Summarize output from qp pipeline

## Usage

```
qp_summarize(x)  
  
## S3 method for class 'data.frame'  
qp_summarize(x)  
  
## S3 method for class 'list'  
qp_summarize(x)
```

## Arguments

x A `data.frame` or a `list` containing a `data.frame` named `qp`

## Value

A `tibble` with the sample name, `sample_type`, and the mean of its predicted concentration (`.pred_conc_mean`)

---

qp\_tidy

*Read in and wrangle protein quantification data*

---

## Description

Read in and wrangle protein quantification data

## Usage

```
qp_tidy(x, ...)

## S3 method for class 'character'
qp_tidy(x, ...)

## S3 method for class 'spectramax'
qp_tidy(
  x,
  replicate_orientation = c("h", "v"),
  n_standards = 7,
  n_replicates = 3,
  wavelength = 562,
  ...
)

## S3 method for class 'gp'
qp_tidy(x, ...)

## Default S3 method:
qp_tidy(x, ...)
```

## Arguments

<code>x</code>	A <code>gp</code> , <code>data.frame/tibble</code> , <code>spectramax</code> , or character path to a raw SPECTRAmax .xls(x)/.txt
<code>...</code>	Arguments passed to relevant methods.
<code>replicate_orientation</code>	Character. Specified the direction the <i>replicates</i> lie, not the direction the samples flow (which will be perpendicular to <code>replicate_orientation</code> ).
<code>n_standards</code>	Numeric. The number of different concentrations of standards. Does not include replicates.
<code>n_replicates</code>	Numeric. The number of replicates per sample.
<code>wavelength</code>	Numeric. For SPECTRAmax files and objects, the wavelength measured. Otherwise, ignored.

## Details

`qp` assumes that if you read in data not in a `spectramax` file or object, you probably have a custom workflow in mind - therefore, tidying will be minimal and mostly focused on checking for validity.

## Value

a `data.frame`

**Examples**

```
data <- system.file("extdata", "absorbances.txt", package = "qp")
qp_tidy(data)
```

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